Vitamin B6 and Pyridoxine Glucoside Content of Wheat and Wheat Flours

D. A. SAMPSON,^{1,2} Q.-B. WEN,³ and K. LORENZ¹

ABSTRACT

Analysis of different wheats and flours for vitamin B6 using water extracts and reverse-phase high-performance liquid chromatography revealed significant variation in content of pyridoxine (PN) and pyridoxine glucoside (PNG), a glycosidic adduct of B6 with partial human bioavailability found in many plant foods. Other vitameric forms of vitamin B6 were not detected. In 22 American and Canadian wheats, total vitamin B6 (the sum of PN plus PNG observed in samples) content varied over a threefold range of 3.4 to 11.1 nmol/g. PNG accounted for a significant portion of the variation (1.9–8.8 nmol/g), with less variation

The U.S. Food and Nutrition Board suggested in the 1989 Recommended Daily Allowances (RDA) (National Research Council 1989) that whole-wheat foods are a good source of vitamin B6 in the American diet. However, a significant and variable fraction of vitamin B6 in plant foods, including wheat, is present as pyridoxine glucoside (5'-O-(B-D-glucopyranosyl)pyridoxine) (PNG) (Gregory and Ink 1987), which has partial bioavailability for humans (Gregory et al 1991). Many Americans consume vitamin B6 at levels significantly less than the current RDA (Kant and Block 1990).

Vitamin B6 includes a group of 3-hydroxy-2-methyl-pyridine derivatives that can exist in the C-4' position as an alcohol (pyridoxine) (PN), an aldehyde (pyridoxal) (PL) and an amine (pyridoxamine) (PM). These vitameric forms can be phosphorylated or nonphosphorylated. The available data suggest that PN predominates in most plant foods (Van Schoonhoven et al 1994). However, most of the reports in the literature have used a microbiological procedure involving an acid hydrolysis step during sample extraction (Toepfer and Lehmann 1961). This procedure converts any PNG in the sample into PN (Gregory 1988). Therefore, in much of the literature about vitamin B6 in plant foods, PNG present in the food may be reported as PN.

We recently reported an high-performance liquid chromatography (HPLC) method capable of measuring vitamin B6 and PNG in wheat (Sampson et al 1995). Although the method does not detect quantitatively minor forms of vitamin B6 such as polyglycosylated adducts (Gregory and Sartain 1991), this method is simple, quick and reliable, and does detect the major forms of vitamin B6 in wheat, including PN and PNG. Our initial analyses of vitamin B6 in several wheat cultivars obtained with this method showed that 36–81% of vitamin B6 in several wheat cultivars was present as PNG (Sampson et al 1995). We have now used this method to investigate the vitamin B6 content of American and Canadian wheats and wheat flours.

Our objectives in this study were: 1) to evaluate the vitamin B6 content, including PNG, of various cultivars of American and Canadian wheat; 2) to determine whether differences in these

²Corresponding author. E-mail: sampson@cahs.colostate.edu.

³South China University of Technology, Guang Zhou, People's Republic of China.

Publication no. C-1996-1007-05R. © 1996 American Association of Cereal Chemists, Inc. vitamers occurred between American and Canadian strains and between hard vs. soft wheats; 3) to compare wheats grown in Colorado to those from other American and Canadian locations; and 4) to determine the extent of loss of these compounds associated with commercial milling. Our interest in American vs. Canadian and Colorado vs. other wheats stemmed from the importance of wheat to Colorado agriculture.

in PN (0.9–2.7 nmol/g). Mean values (\pm standard deviations) were 5.0 \pm

1.8 nmol/g for total vitamin B6; 1.7 ± 0.5 nmol/g for PN; and 3.4 ± 1.8 nmol/g for PNG. American hard wheats contained significantly more

vitamin B6, mostly as PNG, compared to Canadian hard wheats. The

same pattern was seen for Colorado hard wheats compared to others. Hard wheats contained more vitamin B6 than did soft wheats. Flours

milled from 16 of the cultivars contained ≈75% less B6 than did the cor-

MATERIALS AND METHODS

Reagents

responding wheats.

Vitamin B6 standards, deoxypyridoxine, and HPLC reagents were obtained from vendors described earlier (Sampson et al 1995).

Sample Preparation

Seventeen hard wheats and 5 soft wheats, as well as flours milled from 16 of these wheats, were obtained from commercial mills in the United States and Canada. Information about milling, patent percentage, and origin of cultivars was obtained from the mills. Wheats were stored at -20° C in 100-ml polycarbonate bottles before analysis. Sample preparation was done under yellow fluorescent lighting. Samples (10 g) were prepared in triplicate as described previously (Sampson et al 1995) using a micro-mill (Lab Apparatus, Cleveland, OH) and homogenizer (Hi-Speed, model 45, Virtis Co., Gardiner, NY). Moisture, protein, and ash contents of the wheats and flours were determined using approved methods (AACC 1995). Protein was expressed as N × 6.25 for grain and as N × 5.7 for flours.

HPLC Equipment and Method

Wheats were analyzed by reverse-phase HPLC using gradient elution and fluorescence detection, as described elsewhere (Sampson et al 1995). Mobile phase A of the binary gradient contained 0.033M phosphoric acid and 0.008M 1-octanesulfonic acid at pH 2.2. Mobile phase B contained 0.033M phosphoric acid and 10% (v/v) acetonitrile at pH 2.2. Fluorescence was enhanced by postcolumn derivitization with sodium bisulfite. The binary gradient involved a linear ramp from 100% A to 100% B in 10 min, 15 min in 100% B, ramp back to 100% A in 4.5 min, and 5.5 min in 100% A. PN and PNG were quantified relative to fluorescence of the internal standard, deoxypyridoxine. Molar fluorescence of PNG was assumed to be equivalent to PN, as shown by Gregory and Ink (1987).

Cereal Chem. 73(6):770-774

¹Department of Food Science and Human Nutrition, Colorado State University Fort Collins, CO 80523.

 TABLE I

 Identification and Proximate Composition of U.S. and Canadian Grains and Flours

			Grain	Flour				
Sample No. or Variety ^a	Wheat Type ^b	Origin ^c	Protein ^d (%)	Extraction (%)	Patent (%)	Flour Type ^e	Protein ^d (%)	Ash ^d (%)
86	WRS	Unknown	13.1	72.0	59.0	В	12.9	0.47
81	WRS	AB	12.5	73.0	80.0	F	12.2	0.47
73	WRS	AB	15.1	76.0		S	14.4	0.48
34	HW	ID	11.7	70.0	100.0	F	10.4	0.43
70	ARS	AB	13.1	75.5		В	12.8	0.48
87	WRS	MB	12.8	75.5	71.5	В	12.5	0.50
39	HS	ID	12.8		71.6	М	11.6	0.42
83	WRS	Unknown	13.1	73.0	52.0	F	11.7	0.38
40	HS	ND	13.6	73.0	98.0	В	12.6	0.51
11	HW	MN	10.9	74.1	96.8	F	9.9	0.41
Colano	HS	CO	15.8					
42	HS	ND	14.6	75.0	100.0	н	13.8	0.54
Waldron	HW	CO	12.8					
46	HS	MN	15.0	74.5	100.0	н	13.9	0.58
Vona	HW	CO	10.6					
Scout	HW	CO	12.0			• • •		
TAM 107	HW	CO	10.7					
79	OSW	ON	10.1	70.0	100.0	S	9.1	0.50
76	OSW	ON	10.4	75.0	100.0	С	8.1	0.36
33	SW	ID	10.2	75.7	50.3	С	7.9	0.35
56	S	WA	9.6	70.5	58.5	С	7.7	0.38
31	SRW	TN	9.2	75.0	82.0	F	nd	nd

^a Data in Tables I, II, and III are presented in the same order to facilitate comparison.

^b HW = hard red winter; SW = soft white; SRW = soft red winter; HS = hard red spring; ARS = Alberta red spring; OSW = Ontario soft winter; WRS = western red spring.

^c MN = Minnesota; ID = Idaho; ND = North Dakota; WA = Washington; TN = Tennessee; AB = Alberta; ON = Ontario; MB = Manitoba; CO = Colorado.

^d 14% moisture basis; nd = not determined.

^e F = family; C = cake; M = macaroni; H = hearth bread; B = baker's bread flour; S = cookie, crackers

	Sample				Total B6
Type ^c	Country ^d	Locale ^e	PN	PNG	
Hard wheats					
WRS	CAN	Unknown	0.87 ± 0.04	2.52 ± 0.08	3.39 ± 0.07
WRS	CAN	AB	0.99 ± 0.07	2.63 ± 0.28	3.62 ± 0.21
WRS ^f	CAN	AB	1.36 ± 0.12	2.58 ± 0.07	3.94 ± 0.19
HW	USA	ID	1.31 ± 0.15	2.68 ± 0.10	3.99 ± 0.19
ARS	CAN	AB	1.35 ± 0.12	2.74 ± 0.34	4.09 ± 0.39
WRS	CAN	MB	1.51 ± 0.22	2.63 ± 0.20	4.14 ± 0.40
HSf	USA	ID	1.88 ± 0.06	2.34 ± 0.14	4.22 ± 0.12
WRS	CAN	Unknown	2.66 ± 0.15	1.80 ± 0.08	4.46 ± 0.22
HS	USA	ND	2.70 ± 0.01	1.88 ± 0.23	4.58 ± 0.23
HW ^f	USA	MN	1.23 ± 0.09	4.10 ± 0.20	5.33 ± 0.29
HS	USA	CO	1.68 ± 0.05	3.72 ± 0.19	5.40 ± 0.18
HS	USA	ND	1.92 ± 0.23	3.89 ± 0.52	5.81 ± 0.75
HW	USA	CO	1.86 ± 0.02	4.37 ± 0.26	6.23 ± 0.27
HS	USA	MN	2.38 ± 0.26	4.06 ± 0.13	6.44 ± 0.39
HW	USA	CO	2.10 ± 0.27	4.88 ± 0.40	6.98 ± 0.55
HW	USA	CO	1.69 ± 0.02	5.43 ± 0.30	7.12 ± 0.29
HW	USA	CO	2.34 ± 0.22	8.76 ± 0.34	11.10 ± 0.56
Soft wheats					
OSW	CAN	ON	1.10 ± 0.04	1.97 ± 0.19	3.07 ± 0.17
OSW	CAN	ON	1.36 ± 0.12	1.88 ± 0.02	3.24 ± 0.11
SW	USA	ID	1.39 ± 0.25	3.09 ± 0.10	4.48 ± 0.35
S	USA	WA	1.02 ± 0.14	3.56 ± 0.04	4.58 ± 0.18
SRW	USA	TN	2.25 ± 0.06	2.34 ± 0.03	4.59 ± 0.09

 TABLE II

 Vitamin B6 Concentrations (nmol/g) in Whole Wheats^{a,b}

^a Values are mean \pm standard deviations for n = 3 replicates, except as indicated. Data are presented in order of increasing total vitamin B6. Data in Tables I, II, and III are presented in the same order to facilitate comparison.

^b PN = pyridoxine; PNG = pyridoxine glucoside; Total B6 = sum of PN + PNG (other vitamins not detected).

^c WRS = western red spring; HW = hard red winter; ARS = Alberta red spring; HS = hard red spring; OSW = Ontario soft winter; SW = soft white; S = soft; SRW = soft red winter.

^d CAN = Canada; USA =United States.

e AB = Alberta; ID = Idaho; MB = Manitoba; ND = North Dakota; MN = Minnesota; CO = Colorado; ON = Ontario; WA = Washington; TN = Tennessee.

f n = 2 replicates.

RESULTS

Sample identification and proximate composition of wholegrains and flours is given in Table I. Percent protein ranged from 9.2 to 15.1% and did not correlate with content of vitamin B6 in the wheats analyzed.

All of the wheats analyzed contained vitamin B6 as PN and PNG, with none of the other B6 vitamers (PL, PLP, PM, PMP) detected by this method. Limits of detection, expressed as signal-to-noise ratio = 3, were $\approx 0.07-0.29$ pmol per 20 µl injection for the different B6 vitamers, corresponding to 0.02-0.07 nmol/g sample. Therefore PL, PLP, PM, and PMP in these samples were present at levels <0.07 nmol/g. Total vitamin B6 will refer to the sum of PN plus PNG observed in samples.

There was significant variation in content of PN and PNG (Table II). Observed total vitamin B6 content varied over a threefold range (3.4-11.1 nmol/g) associated with a smaller variation in PN (0.9-2.7 nmol/g) and a larger variation in PNG (1.9-8.8 nmol/g). The means \pm standard deviations for these ranges were: 5.0 ± 1.8 nmol/g, total vitamin B6; 1.7 ± 0.5 nmol/g, PN; 3.4 ± 1.8 nmol/g, PNG. Most cultivars contained vitamin B6 in a range of 3.4-7.1 nmol/g for total B6 and 1.9-5.4 nmol/g for PNG. One American cultivar (HW, USA, CO) contained significantly more vitamin B6 than other cultivars, with 11.1 nmol/g for total B6 and 8.8 nmol/g for PNG. The contributions of PN and PNG to total vitamin B6 in these wheats varied in lower vs. higher B6-containing cultivars. In cultivars containing total vitamin B6 in the range of 3-5 nmol/g, samples with higher totals had more PN. In contrast, in cultivars containing vitamin B6 in the range of 5-11 nmol/g, samples with higher totals had more PNG.

American wheats (11 cultivars) showed a wide range of total vitamin B6 content (4.0–11.1 nmol/g for total vitamin B6) in contrast to Canadian wheats (six cultivars), which covered a narrower range of 3.4–4.5 nmol/g for total vitamin B6 (Table II). On average, the American cultivars contained 56% more total vitamin B6 than did those from Canada ($6.1 \pm 2.0 \text{ vs. } 3.9 \pm 0.4 \text{ nmol/g}$, respectively). Most of this difference was due to higher PNG content, which averaged 68% higher in the American wheats when compared to Canadian wheats ($4.2 \pm 1.9 \text{ vs. } 2.5 \pm 0.3 \text{ nmol/g}$, respectively). This differences between American and

Canadian wheats remained even when the American cultivar with particularly high total vitamin B6 (HW, USA, CO) was removed from analysis, although the average percent differences for total vitamin B6 and PNG decreased to +44% and +48%, respectively.

The total vitamin B6 and PNG content tended to be higher in hard wheats than in soft wheats (Table II). For hard wheats, the range (and mean \pm standard deviation) were: 3.4–11.1 nmol/g (5.3 \pm 1.9 nmol/g) for total vitamin B6; 0.9–2.7 nmol/g (1.8 \pm 0.6 nmol/g) for PN; and 1.8–8.8 nmol/g (4.0 \pm 1.7 nmol/g) for PNG. The corresponding values for soft wheats were: 3.1–4.6 nmol/g (4.0 \pm 0.8 nmol/g) for total vitamin B6; 1.0–2.2 nmol/g (1.4 \pm 0.5 nmol/g) for PN; and 1.9–3.6 nmol/g (2.6 \pm 0.7 nmol/g) for PNG. Thus, on average, hard wheats contained 32% more total vitamin B6 than did soft wheats, accounted for primarily by 54% more PNG.

Compared to other hard wheats, those from Colorado (five cultivars) contained 61% more total vitamin B6 (7.4 \pm 2.2 vs. 4.6 \pm 0.9 nmol/g) comprised primarily of 80% more PNG (5.4 \pm 2.0 vs. 3.0 \pm 0.8 nmol/g), with little difference between PN content (1.9 \pm 0.3 vs. 1.7 \pm 0.6 nmol/g). Four out of five cultivars with highest total B6 and PNG content were from Colorado. The wheat cultivar containing the highest content of total B6 as well as PNG is one of two most widely grown wheats in Colorado (TAM-107). The other widely grown Colorado wheat (Vona) had similarly high total B6 and PNG (7.0 \pm 0.6 nmol/g for total B6).

Milling whole wheats into flours removed significant amounts of vitamin B6 (Tables III vs. II). For 12 flours prepared from hard wheats and four from soft wheats, the average loss of total vitamin B6 from milling was -74% (4.3 ± 0.9 vs. 1.1 ± 0.4 nmol/g). Percent losses were somewhat higher for PNG (-79%) (2.8 ± 0.8 vs. $0.6\pm$ 0.2 nmol/g) than for PN (-69%) 1.6 ± 0.6 vs. 0.5 ± 0.2 nmol/g). For two cultivars with low patent percent (50-52%), the loss in PN for flour vs. whole wheat (- $81\pm1\%$) was significantly higher than for 12 cultivars with higher patent percent (58-100%) (- $63\pm6\%$).

DISCUSSION

The main conclusion from our data is that wheats from the United States and Canada contain easily extractable vitamin B6 in a range from 3 to 11 nmol/g, and that flours from those wheats contain \approx 1 nmol of total vitamin B6/g. Most reports in the litera-

	Sample				Total B6
Турес	Country ^d	Locale ^e	PN	PNG	
Hard wheats					
WRS	CAN	Unknown	0.34 ± 0.01	0.52 ± 0.05	0.85 ± 0.06
WRS	CAN	AB	0.33 ± 0.02	0.51 ± 0.07	0.84 ± 0.09
WRS ^f	CAN	AB	0.49 ± 0.04	0.42 ± 0.06	0.91 ± 0.08
HW	USA	ID	0.43 ± 0.03	0.71 ± 0.04	1.14 ± 0.06
ARS	CAN	AB	0.49 ± 0.04	0.67 ± 0.01	1.16 ± 0.04
WRS	CAN	MB	0.36 ± 0.01	0.90 ± 0.05	1.26 ± 0.06
HSf	USA	ID	0.82 ± 0.03	0.28 ± 0.04	1.10 ± 0.06
WRS	CAN	Unknown	0.47 ± 0.04	0.53 ± 0.03	1.00 ± 0.07
HS	USA	ND	1.04 ± 0.24	0.77 ± 0.12	1.81 ± 0.29
HW ^f	USA	MN	0.51 ± 0.01	0.38 ± 0.04	0.89 ± 0.04
HS	USA	ND	0.64 ± 0.02	0.96 ± 0.02	1.61 ± 0.04
HS	USA	MN	0.88 ± 0.07	0.70 ± 0.06	1.57 ± 0.02
Soft wheats					
OSW	CAN	ON	0.47 ± 0.05	0.71 ± 0.05	1.18 ± 0.04
OSW	CAN	ON	0.47 ± 0.01	0.33 ± 0.05	0.80 ± 0.05
SW	USA	ID	0.27 ± 0.03	0.12 ± 0.03	0.39 ± 0.06
S	USA	WA	0.43 ± 0.04	0.35 ± 0.03	0.79 ± 0.07

 TABLE III

 Vitamin B6 Concentrations (nmol/g) in Wheat Flours^{a,b}

^a Values are mean \pm standard deviations for n = 3 replicates, except as indicated. Data in Tables I, II, and III are presented in the same order to facilitate comparison.

^b PN = pyridoxine; PNG = pyridoxine glucoside; Total B6 = sum of PN + PNG (other vitamins not detected).

^c WRS = western red spring; HW = hard red winter; ARS = Alberta red spring; HS = hard red spring; OSW = Ontario soft winter; SW = soft white; S = soft.

^d CAN = Canada; USA =United States.

^c AB = Alberta; ID = Idaho; MB = Manitoba; ND = North Dakota; MN = Minnesota; CO = Colorado; ON = Ontario; WA = Washington. ^f n = 2 replicates. ture suggest that whole wheat contains $\approx 23 \text{ nmol/g}$ for vitamin B6, mostly present as PN ($\approx 16 \text{ nmol/g}$), with lesser amounts of PL and PM (2–4 nmol/g, each) (Toepfer and Lehmann 1961, Polansky et al 1964, Toepfer and Polansky 1970, Toepfer et al 1972, Michaela and Lorenz 1976, Keagy et al 1980, Vanderslice et al 1984). Whole wheat flour has been reported to contain ≈ 19 nmol/g for vitamin B6, present primarily as PN (15 nmol/g) (Toepfer and Polansky 1964, 1970; Michaela and Lorenz 1976; Kabir et al 1983). Two reports show that white flour contains much lower amounts of vitamin B6 (2–4 nmol/g) (Michaela and Lorenz 1976, Keagy et al 1980).

That our vitamin B6 values for wheat are lower than in earlier reports raises the possibility of incomplete extraction using our method. However, we reported previously that our method gives complete extraction of PN, measured using exogenous PN spikes. We have assumed that PNG recovery is similarly high. Two other laboratories have reported that PNG extraction from plant foods is as high as that for PN (Gregory and Feldstein 1985, Tadera et al 1986, Gregory and Ink 1987, Tadera and Naka 1991). Gregory (Gregory 1988) has pointed out that recovery of exogenous spikes added during sample preparation does not necessarily reflect recovery of endogenous compounds in foods. Resolution of discrepancy between our wheat B6 values and those in earlier reports will await future studies measuring recovery of intrinsically labeled B6 vitamers (Gregory 1988).

A second conclusion from our data is that a significant fraction (average of 68%) of vitamin B6 in wheat is present as PNG. There is a paucity of reports documenting PNG content of whole wheat, and only one report dealing with PNG in wheat flour (Kabir et al 1983). Total B6 and PN content of wheat bran (\approx 60 and 23 nmol/g, respectively) is reported to be three-fold higher than in wheat or flour (Michaela and Lorenz 1976, Kabir et al 1983, Tadera et al 1986, Gregory and Sartain 1991). In two of these reports (Tadera et al 1986, Gregory and Sartain 1991), PNG, measured using HPLC, comprised a significant portion of total vitamin B6 (\approx 40%) (Gregory and Sartain 1991). These observations of PNG in wheat bran are consistent with our data that a significant portion of wheat B6 is PNG (Toepfer and Lehmann 1961, Polansky et al 1964, Toepfer and Polansky 1964, 1970, Toepfer et al 1972, Michaela and Lorenz 1976, Kabir et al 1983).

PNG has bioavailability in human subjects of $\approx 58\%$ (Gregory et al 1991) compared to PN, so that the amount of available B6 in wheat will be significantly lower than the total amount if PNG were present in significant amounts. This could be potentially important for vitamin B6 nutritional status in individuals who habitually consume large quantities of unfortified whole wheat products, especially if their diet provides marginal amounts of B6 relative to the RDA. Food intake surveys have shown repeatedly that many Americans consume inadequate amounts of vitamin B6.

Glycosylated forms of vitamin B6 other than PNG occur in wheat, but at levels much lower than for PNG itself (Tadera et al 1988, Gregory and Sartain 1991). Our method's relative quickness and simplicity precluded our obtaining information in the wheats we analyzed about these other minor glycosylated forms of B6.

The source of variation in wheat PN and PNG reported here may be storage time and condition, seasonal variation, and geographic variation.

The variation in vitamin B6 content for flour vs. whole wheat confirms earlier reports (Schroeder 1971) showing that milling removes a significant portion (\approx 75%) of vitamin B6. This observation emphasizes that milling significantly decreases the nutrient density of vitamin B6 in wheats.

In conclusion, our results indicate that wheat contains variable amounts of vitamin B6 with significant amounts accounted for by PNG. Our observations about vitamin B6 in American vs. Canadian wheats, Colorado vs. other wheats, and hard vs. soft wheats emphasize the larger point that vitamin B6 content of wheats varies considerably between cultivars. Using water and metaphosphoric acid extraction, we have observed vitamin B6 content in wheats that are lower than in earlier reports using standard acid extraction procedures (AOAC). Resolution of this numerical disparity awaits a future study focused on comparison of extraction methods. Our primary interest and conclusion is that vitamin B6 concentration varies significantly within wheats, and that PNG accounts for a significant percentage of vitamin B6 in wheats.

ACKNOWLEDGMENTS

We thank Bridget Farrell, Peter Lorenz, Brenda Grieser and Laurie Wenzel for technical assistance with sample extraction.

LITERATURE CITED

- AMERICAN ASSOCIATION OF CEREAL CHEMISTS. 1995. Approved Methods of the AACC, 9th ed. Method 8-01, approved April 1961, revised October 1976 and October 1981, reviewed October 1994; Method 30-20, approved April 1961, revised October 1975 and October 1982, reviewed October 1994; Method 44-15A, approved October 1975, revised October 1981 and October 1994; Method 46-12, approved October 1976, revised October 1986, reviewed October 1994; Method 86-31, approved April 1968, revised October 1981, reviewed October 1994. The Association: St. Paul, MN.
- GREGORY, J. F. 1988. Methods for determination of vitamin B6 in foods and other biological materials: A critical review. J. Food Compos. Anal. 1:105-123.
- GREGORY, J. F., and FELDSTEIN, D. 1985. Determination of vitamin B-6 in foods and other biological materials by paired-ion high-performance liquid chromatography. J. Agric. Food Chem. 33:359-363.
- GREGORY, J. F., and INK, S. L. 1987. Identification and quantification of pyridoxine-beta-glucoside as a major form of vitamin B-6 in plantderived foods. J. Agric. Food Chem. 35:76-82.
- GREGORY, J. F., and SARTAIN, D. B. 1991. Improved chromatographic determination of free and glycosylated forms of vitamin B6 in foods. J. Agric. Food Chem. 39:899-905.
- GREGORY, J. F., TRUMBO, P. R., BAILEY, L. B., TOTH, J. P., BAUMGARTNER, T. G., and CERDA, J. J. 1991. Bioavailability of pyridoxine-5'-B-D-glucoside determined in humans by stable-isotopic methods. J. Nutr. 121:177-186.
- KABIR, H., LEKLEM, J. E., and MILLER, L. T. 1983. Measurement of glycosylated vitamin B6 in foods. J. Food Sci. 48:1422-1425.
- KANT, A. K., and BLOCK, G. 1990. Dietary vitamin B-6 intake and food sources in the U.S. population: NHANES II, 1976-1980. Am. J. Clin. Nutr. 52:707-716.
- KEAGY, P. M., BORENSTEIN, B., RANUM, P., CONNOR, M. A., LORENZ, K., HOBBS, W. E., HILL, G., BACHMAN, A. L., BOYD, W. A., and KULP, K. 1980. Natural levels of nutrients in commercially milled wheat flours. II. Vitamin analysis. Cereal Chem. 57:59-65.
- MICHAELA, P., and LORENZ, K. 1976. The vitamins of triticale, wheat and rye. Cereal Chem. 53:853-861.
- NATIONAL RESEARCH COUNCIL. 1989. Recommended Dietary Allowances, 10th ed. National Academy of Sciences: Washington DC.
- POLANSKY, M. M., MURPHY, E. W., and TOEPFER, W. E. 1964. Components of vitamin B6 in grains and cereal products. J. Assoc. Off. Agric. Chem. 47:750-753.
- SAMPSON, D. A., EOFF, L. A., YAN, X. L., and LORENZ, K. 1995. Analysis of free and glycosylated vitamin B6 in wheat by high-performance liquid chromatography. Cereal Chem. 72:217-221.
- SCHROEDER, H. A. 1971. Losses of vitamins and trace minerals resulting from processing and preservation of foods. Am. J. Clin. Nutr. 24:562-573.
- TADERA, K., and NAKA, Y. 1991. Isocratic paired-ion high-performance liquid chromatographic method to determine B6 vitamers and pyridoxine glucoside in foods. Agric. Biol. Chem. 55:563-564.
- TADERA, K., KANEKO, T., and YAGI, F. 1986. Evidence for the occurrence and distribution of a new type of vitamin B6 conjugate in plant foods. Agric. Biol. Chem. 50:2933-2934.
- TADERA, K., KANEKO, T., and YAGI, F. 1988. Isolation and structural elucidation of three new pyridoxine-glycosides in rice bran. J. Nutr. Sci. Vitaminol. 34:167-177.
- TOEPFER, E. W., and LEHMANN, J. 1961. Procedure for chromatographic separation and microbiological assay of pyridoxine, pyridoxal, and pyridoxamine in food extracts. J. AOAC 44:426-430.

- TOEPFER, E. W., and POLANSKY, M. M. 1964. Recent developments in the analysis for vitamin B6 in foods. Pages 825-832 in: Vitamins and Hormones. Vol. 22. R. S. Harris, I. G. Wool, and J. A. Loraine, eds. Academic Press: New York.
- TOEPFER, E. W., and POLANSKY, M. M. 1970. Microbiological assay of vitamin B6 and its components. J. Assoc. Off. Agric. Chem. 53:546-550.
- TOEPFER, E. W., POLANSKY, M. M., EHEART, J. F., SLOVER, H. T., MORRIS, E. R., HEPBURN, F. N., and QUACKENBUSH, F. W. 1972. Nutrient composition of selected wheats and wheat products. XI.

Summary. Cereal Chem. 49:173-186.

- VAN SCHOONHOVEN, J., SCHRIJVER, J., VAN DEN BERG, H., and HAENEN, G. R. M. M. 1994. Reliable and sensitive high-performance liquid chromatographic method with fluorometric detection for the analysis of vitamin B-6 in foods and feeds. J. Agric. Food Chem. 42:1475-1480.
- VANDERSLICE, J. T., BROWNLEE, S. R., and CORTISSOZ, M. E. 1984. Liquid chromatographic determination of vitamin B-6 in foods. J. Assoc. Off. Anal. Chem. 67:999-1006.

[Received December 7, 1995. Accepted June 13, 1996.]